



细胞、细菌透射电镜实验报告

一、实验器材及试剂

1、实验器材

名称	厂家	型号
超薄切片机	Leica	Leica UC7
钻石切片刀	Daitome	Ultra 45°
透射电子显微镜	HITACHI	HT7800/HT7700
150 目方华膜铜网		

2、主要实验试剂

试剂	厂家	货号
电镜固定液	Servicebio	G1102
无水乙醇	国药集团化学试剂有限公司	100092183
丙酮	国药集团化学试剂有限公司	10000418
812 包埋剂	SPI	90529-77-4
锇酸	Ted Pella Inc	

二、透射电镜制片步骤

- 取材固定：**离心收集细胞或细菌沉淀，要求沉淀最少绿豆大小。去培养基加入电镜固定液 4℃重悬混匀固定 2-4h，4℃固定保存及运输。
- 琼脂预包埋：**细胞或细菌用离心机离心，弃上清加入 0.1M 磷酸缓冲液 PB（PH7.4），混匀漂洗 3min 后再离心，重复洗涤 3 次。提前加热溶解制备 1%琼脂糖溶液，稍冷却后加入 EP 管内，在琼脂糖凝固之前将沉淀用镊子挑起悬浮包裹于琼脂糖内。
- 后固定：**0.1M 磷酸缓冲液 PB（PH7.4）配制的 1%锇酸避光室温固定 2h。0.1M 磷酸缓冲液 PB（PH7.4）漂洗 3 次，每次 15min。
- 室温脱水：**组织依次入 30%-50%-70%-80%-95%-100%-100%酒精上行脱水每次 20min，100%丙酮两次，每次 15min。
- 渗透包埋：**丙酮：812 包埋剂=1：1 37℃ 2-4h， 丙酮：812 包埋剂=1：2 37℃渗透过夜，纯 812 包埋剂 37℃ 5-8h。将纯 812 包埋剂倒入包埋板，将样品插入包埋板后 37℃烤箱过夜。
- 聚合：**包埋板放于 60℃烤箱聚合 48h，取出树脂块备用。
- 超薄切片：**树脂块于超薄切片机 60-80nm 超薄切片，150 目方华膜铜网捞片。



武汉赛维尔生物科技有限公司

Wuhan Servicebio Technology Co., Ltd.

8、染色：铜网于 2%醋酸铀饱和酒精溶液避光染色 8min；70%酒精清洗 3 次；超纯水清洗 3 次；2.6%枸橼酸铅溶液避二氧化碳染色 8min；超纯水清洗 3 次，滤纸稍吸干。铜网切片放入铜网盒内室温干燥过夜。

9、透射电子显微镜下观察，采集图像分析。

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TEM Staining Report for Cells and Bacteria

1 Apparatus and reagents

1.1 Major apparatus

Name	Producer	Model
Ultra microtome	Leica	Leica UC7
Diamond slicer	Daitome	Ultra 45°
Transmission Electron Microscope	HITACHI	HT7800/HT7700
150 meshes cuprum grids with formvar film		

1.2 Major reagents

Name	Producer	Code
Fixative for TEM	Servicebio	G1102
Ethanol	Sinaopharm Group Chemical Reagent Co. LTD	100092183
Aceton	Sinaopharm Group Chemical Reagent Co. LTD	10000418
EMBed 812	SPI	90529-77-4
OsO ₄	Ted Pella Inc	

2 Procedure

2.1 Harvest samples and fixation: Collect cells or bacteria precipitation after centrifuge, requiring the precipitation should be at least mung beans size. The TEM fixative was added to the tube and let the precipitation re-suspended in the fixative, and then fixed at 4 °C for preservation and transportation.

2.2 Agarose pre-embedding: The fixed cells and bacteria were centrifuged. The 0.1 M PB (pH 7.4) was added into the tube after supernatant was discarded, and then the precipitation was re-suspended and washed in PB for 3min. This washing step was repeated for 3 times. The 1% agarose solution was prepared by heating and dissolving in advance. After being cooled, the agarose solution was added into the EP tube. Before agarose solidification, the precipitation was suspended with forceps and wrapped in the agarose.

2.3 Post-fix: Agarose blocks with samples avoid light post fixed with 1% OsO₄ in 0.1 M PB (pH 7.4) for 2 h at room temperature. After remove OsO₄, the tissues are rinsed in 0.1 M PB (pH 7.4)



for 3 times, 15 min each.

2.1 Dehydrate at room temperature as followed:

30% ethanol for 20 min;
50% ethanol for 20 min;
70% ethanol for 20 min;
80% ethanol for 20 min;
95% ethanol for 20 min;
Two changes of 100% ethanol for 20 min;
Finally two changes of acetone for 15 min.

2.4 Resin penetration and embedding as followed:

Acetone: EMBED 812=1:1 for 2-4 h at 37°C;
Acetone: EMBED 812=1:2 overnight at 37°C;
pure EMBED 812 for 5-8 h at 37°C;

Pour the pure EMBED 812 into the embedding models and insert the tissues into the pure EMBED 812, and then keep in 37°C oven overnight.

2.5 Polymerization: The embedding models with resin and samples were moved into 65°C oven to polymerize for more than 48h. And then the resin blocks were taken out from the embedding models for standby application at room temperature.

2.6 Ultrathin section: The resin blocks were cut to 60-80nm thin on the ultra microtome, and the tissues were fished out onto the 150 meshes cuprum grids with formvar film.

2.7 Staining: 2% uranium acetate saturated alcohol solution avoid light staining for 8 min, rinsed in 70% ethanol for 3 times and then rinsed in ultra pure water for 3 times. 2.6% Lead citrate avoid CO₂ staining for 8 min, and then rinsed with ultra pure water for 3 times. After dried by the filter paper, the cuprum grids were put into the grids board and dried overnight at room temperature.

2.8 Observation and images capture: The cuprum grids are observed under TEM and take images.