masson 染色实验报告

一、实验器材及试剂

1、 实验器材

名称	厂家	型号
脱水机	DIAPATH	Donatello
包埋机	武汉俊杰电子有限公司	JB-P5
病理切片机	上海徕卡仪器有限公司	RM2016
冻台	武汉俊杰电子有限公司	JB-L5
组织摊片机	浙江省金华市科迪仪器设备有限公司	KD-P
烤箱	天津市莱玻瑞仪器设备有限公司	GFL-230
载玻片	Servicebio	G6004
正置光学显微镜	日本尼康	NIKON ECLIPSE E100
成像系统	日本尼康	NIKON DS-U3

2、 主要实验试剂

试剂名称	厂家	货号
无水乙醇	国药集团化学试剂有限公司	100092683
二甲苯	国药集团化学试剂有限公司	10023418
masson 染液套装	Servicebio	G1006
中性树胶	国药集团化学试剂有限公司	10004160

二、实验步骤

1、石蜡切片脱蜡至水：依次将切片放入二甲苯I20min-二甲苯II20min-无水乙醇I5min-无水乙醇II5min-75%酒精 5min，自来水洗。

2、切片浸入 Masson A 液中浸泡过夜，自来水洗。

3、切片入 Masson B 液及 Masson C 液等比混合的染液，浸染 1min，自来水洗，1%盐酸酒精分化，自来水洗。

4、切片入 Masson D 液浸染 6 min，自来水漂洗。

5、Masson E 液浸染 1 min。

6、不水洗，稍沥干直接入 Masson F 液染 2-30s。

7、切片用 1%冰醋酸漂洗分化，两缸无水乙醇脱水。

8、透明封片：切片放入第三缸无水乙醇 5min，二甲苯 5min 透明，中性树胶封片。

9、显微镜镜检，图像采集分析。

三、结果判读：

胶原纤维呈蓝色；肌纤维、纤维素和红细胞呈红色。

四、注意事项：

1、Masson A 液染完后水洗要稍快，切片洗干净即可。

2、根据切片的数量对混合的染液进行更换，也可以现配现用；

3、Masson D 液的染色程度要控制好，胶原部分不能是红色或者太红，会影响后续 Masson F 液的着色；

4、冰醋酸分化过程中，若是分化过度，胶原蓝色太浅，若分化不足，易与红色叠加呈紫蓝色。

Masson Staining Report

1 Apparatus and Reagents

1.1 Major apparatus

Name	Producer	Model
Dehydrator	DIAPATH	Donatello
Embedding machine	Wuhan Junjie Electronics Co., Ltd	JB-P5
Pathology slicer	Leica	RM2016
Frozen platform	Wuhan Junjie Electronics Co., Ltd	JB-L5
Organizer	KEDEE	KD-P
oven	Labotery	GFL-230
Glass slide	Servicebio	G6004
Upright optical microscope	Nikon	NIKON ECLIPSE E100
Imaging system	Nikon	NIKON DS-U3

1.2 Major reagents

Name	Producer	Code
Ethanol	SCRC	100092683
Xylene	SCRC	10023418
Masson dye solution set	Servicebio	G1006
Neutral gum	SCRC	10004160

2 Procedure

2.1 Dewaxing as followed:

Xylene I for 20 min;
Xylene II for 20 min;
100% ethanol I for 5 min;
100% ethanol II for 5 min;
75% ethanol for 5 min;
Rinsing with tap water ;

2.2 Soak the slices in Masson A overnight, rinse with tap water.

2.3 Masson B, Masson C were prepared into Masson solution according to the ratio of 1:1. Then stain with Masson solution for 1 min, rinse with tap water. And differentiation it with 1% hydrochloric acid alcohol, rinse with tap water.

2.4 Soak the slices in Masson D for 6 min, rinse with tap water; Masson E for 1 min; Masson F for 2-30s.

2.5 Rinse the slices with 1% glacial acetic acid and then dehydration with two cup of anhydrous ethanol.

2.6 100% ethanol for 5 min; Xylene for 5 min; Finally seal with neutral gum.

2.7 Observe with microscope inspection, image acquisition and analysis.

3 Results

Color	Result
Blue	Collagen fibers
Red	Muscle fibers; Cellulose; Erythrocyte

4 Precautions

4.1 After treat the slics with Masson A , raine with the water should be quikly. It is sufficient to just wash the slices clean.

4.2 Replace the mixed dye solution according to the number of slices. And it can also be formulated when you need.

4.3 The staining degree of Masson D solution should be well controlled. The collagen part cannot be red, which will affect the subsequent coloration of Masson F solution;

4.4 During the differentiation of glacial acetic acid, if it is over-differentiated, the collagen blue will be too light, if it is insufficiently differentiated, it will be easily superimposed with red to become purple-blue .